



NFI BULLETIN

Bulletin of the Nutrition Foundation of India

Volume 28 Number 1

January 2007

Role of Muscle in Fat/Glucose Metabolism and Insulin Resistance

AV Kurpad, S Ramprakash, P Pai, M Vaz

Insulin resistance (IR) and obesity, which are part of the metabolic syndrome, are among the important antecedents to the development of non communicable diseases of aging (NCD) such as diabetes and coronary heart disease (CHD), which are increasing rapidly in India^{1,2}. At a metabolic level, IR translates into an inefficient insulin function in end organs such as skeletal muscle, liver and adipocytes. IR is strongly associated with measures of overall obesity and the location of body fat in Indians^{2,3}. The greater pre-dilection for Indians to develop IR at low body weight may, in part, be due to their relatively greater adiposity at a lower body mass index (BMI) compared with other ethnic groups, both within and outside Asia³⁻⁷. However, while total body fat, or its distribution are important determinants of IR, it is important to independently assess the role of skeletal muscle. There are several considerations in this assessment, which include the actual muscle mass, fiber type, the accumulation of fat within muscle, and muscle metabolism in response to dietary stimuli and physical activity.

BMI, Muscle Mass and Fiber Type

A simple framework for the assessment of the effect of skeletal muscle on IR would be one which considered the effect of total body muscle mass, which has an independent effect on insulin sensitivity and glucose disposal⁸⁻¹¹. It may be that Indians have a lower total body muscle mass, since a large percentage of Indians have a low to

normal BMI¹², and studies on Indians or Asians (adult and pre-pubertal children) have indicated that they have a relatively low muscle mass as well, in comparison to Caucasian and African Americans^{6,13,14}. It is not clear if the cause for this is genetic, or related to environmental factors. In Indians, a low muscle mass can be further worsened by chronic undernutrition¹⁵. Several studies in Indians within a BMI range of 23 to 25 kg/m² have shown lower insulin sensitivity in these subjects compared to age and BMI matched Caucasian counterparts^{3,16,17}. While an inverse relation between glucose disposal rate or the insulin sensitivity index and visceral, subcutaneous and total fat has also been observed for both Indians and Caucasians^{16,17}, the curve for each of these age and BMI matched groups was different, with different intercepts. The latter point would suggest that factors other than the body fat and its location are operative in determining insulin sensitivity. Indeed, the insulin sensitivity index of Indians remained lower than that of age and BMI matched Caucasians even after adjustment for total body fat and truncal skinfold thickness¹⁶. Habitual physical activity levels and muscle mass are two factors that may explain the observed differences between Indians and Caucasians. Although muscle mass was not measured in the afore-mentioned studies, Raji et al¹⁷ described their Indian subjects as being much less active than their Caucasian counterparts. The need for data on IR in subjects with similar muscle mass but different physical

activity levels is evident.

However, a simple framework of a low total body muscle mass and a high risk of IR is insufficient when one considers the complexity of skeletal muscle fiber type, fat accumulation within muscle and the large number of intermediary regulatory molecules that operate within this tissue. The heterogeneity of muscle within the body is also relevant as glucose clearance is different across different muscle beds in the upper and lower limb of Type 2 diabetic patients¹⁸. Comparison studies between African American and Caucasian women have revealed that the former had lower insulin sensitivity in spite of having a higher muscle volume¹⁹. Indeed, the skeletal muscle volume accounted for the difference in insulin response between the groups. The reason for the higher muscle volume could conceivably be due to fat accumulation within the whole muscle. Intramyocellular lipid (IMCL) has been described earlier²⁰ and it has been speculated that the insulin resistance in Indians may be partly due to this alteration in muscle². In addition, fat can be deposited between muscle fibers as intermuscular adipose tissue (IMAT)²¹, and this has also been

CONTENTS

● Role of Muscle in Fat/Glucose Metabolism and Insulin Resistance – AV Kurpad, S Ramprakash, P Pai, M Vaz	1
● Foundation News	6
● Nutrition News	6
● The “Dual Nutrition Burden”- Need for a Balanced Response – C.Gopalan	7

shown to be independently related to IR in African American women, even though it only weakly attenuates the significant difference in IR between African American and Caucasian women¹⁹. While this suggests that simple muscle bulk measurements may not suffice any longer, it must be pointed out that all the foregoing assessments were made in women with a BMI near 30 kg/M², and that in lower BMI Indians with a possible low insulin sensitivity, the total muscle mass may still have a negative association with IR. Furthermore, this also underscores the necessity for functional measurements to assess the role of muscle in IR and NCD. Muscle strength, which is known to decline with age²², has been shown to predict all cause mortality independent of BMI in middle aged American Japanese men²³. Cross-sectional studies on young South Indian men with chronic undernutrition have shown that they have a reduced forearm skeletal muscle mass and hand grip strength²⁴. Fiber types in muscle are relevant and there is a significant reduction in the proportion of slow oxidative fibers and a significant increase in the proportion of fast glycolytic fibers in muscle from type 2 diabetic patients, contributing to the reduced oxidative capacity of skeletal muscles of type 2 diabetic patients²⁵. This is relevant as described below.

Muscle and Insulin Resistance

Normal living is a state of alternating feeding and fasting. During the former, muscle stores are replenished, and during the latter stores are used up, and skeletal muscle adapts well to these fluctuations by increasing its reliance upon fat oxidation²⁶. This flexibility of fuel selection allows for sparing muscle glycogen during exercise and reducing the uptake of plasma glucose. The decrease in muscle glucose uptake is accompanied by a reduction in oxidative glucose disposal with little change in non-oxidative glucose disposal^{27,28}. The oxidative disposal of glucose is dependent on suppression of pyruvate dehydrogenase complex (PDC) activity, which in turn is inhibited by phosphorylation of its E1 component by pyruvate dehydrogenase kinase (PDK). In humans, 48 hours of starvation has been shown to downregulate the expression of hexokinase II activity five-fold whereas PDK expression was upregulated four-fold²⁹. The important finding in this

study was that starvation did not affect the expression of genes involved in fat metabolism, and that the change in substrate oxidation was dependent on downregulating the enzymes involved in oxidative glucose metabolism²⁹. An interesting study on expression profiling of muscle mRNA after weight loss in morbidly obese women was able to map three important genes into the same network that was responsive to weight loss³⁰. These were growth factor receptor-bound protein 14 (GRB14), glycerol-3-phosphate dehydrogenase 1 (GPD1), and growth differentiation factor 8 (GDF8; myostatin), which significantly reduced by about half or more, after weight loss³⁰. While GRB14 inhibits the action of insulin and GPD1 is associated with increased triglyceride synthesis, GDF8 or myostatin negatively regulates muscle mass³¹. Expression profiling may be a relatively blunt tool especially in studies of a small number of subjects; however, this study raises the interesting linkage of a gene for muscle mass with those related to insulin sensitivity.

Skeletal muscle, in a state of IR, has an increased muscle glucose oxidation under basal conditions³². This is in contrast to what might be expected due to the Randle effect³³, which hypothesizes that increased fat availability and oxidation in muscle would lead to a reduced glucose oxidation via the effects of acetyl-CoA on pyruvate dehydrogenase. In essence, therefore, IR could produce a state of "metabolic inflexibility", which would contribute to the accumulation of triglyceride within the myocyte³⁴. This has also been confirmed in human muscle culture experiments where the capacity for fat oxidation by skeletal muscle was increased in subjects with increased insulin sensitivity, leanness, and aerobic fitness³⁵. This is consistent with observations in whole body experimentation, where fasting skeletal muscle in individuals with type 2 diabetes and obesity showed (through measurements of the respiratory quotient across the lower limb tissue bed) increased levels of carbohydrate oxidation in type 2 diabetes and in obesity³⁶, and this low ratio of (fasting) fat to carbohydrate oxidation is thought to be predictive of weight gain³⁷. A note of caution however is to be sounded about the universal applicability of these findings: while muscle triglyceride (as IMCL) was related to insulin sensitivity in Europeans, it was

not similarly related in South Asians, suggesting that other factors may be important in this population³⁸.

Fatty Acids and Muscle Function

An increase in circulating free fatty acids (FFA) due to increased fat mass can precipitate a state of IR through a variety of mechanisms and there may be no single or common defect that can be pinpointed. Some of these are described below. Skeletal muscle glucose-6-phosphate levels decrease during glucose clamp studies when circulating FFA levels are elevated by lipid/heparin infusion³⁹, along with a lower intracellular free glucose levels indicating that FFA directly inhibit glucose transport and phosphorylation. Lipid infusions lead to a decrease in both insulin-stimulated tyrosine phosphorylation of insulin receptor substrate (IRS)-1 as well as activation of phosphoinositide-3 kinase 3 (PI3K) in skeletal muscle⁴⁰. These are important downstream signaling mechanisms for glucose transporter translocation, although their role is still to be fully elucidated. Palmitic acid (C16:0) has been shown to be a potent inhibitor of insulin-induced glucose transport⁴¹, through inhibition of the serine/threonine kinase Akt, also known as protein kinase B (PKB), which acts downstream of IRS-1-coupled PI3K. In mice skeletal muscle cells, palmitate is a potent inhibitor (by 50%) of the insulin receptor signal based phosphorylation of PKB/Akt, which in turn is responsible for translocation of glucose transporters (GLUT 4)⁴¹. Both LCFA-CoAs and DAG can activate protein kinase C (PKC), especially atypical PKC isozymes such as PKC θ ⁴², which can serine phosphorylate IRS-1 and impair its ability to associate with the insulin receptor⁴³, thereby interfering with PI3K activation. Recent studies have also implicated c-jun amino-terminal kinase (JNK) as a mediator of FFA induced insulin resistance⁴⁴. JNK is a member of the p42/p44 mitogen-activated protein (MAP) kinase superfamily which consists of downstream signaling molecules of the insulin pathway. JNK is activated by FFA⁴⁵ and influences gene transcription by transcription factors such as c-jun and ATF 2 (activating transcription factor 2), suggesting a role for this interaction in impaired insulin signaling⁴⁶. Another mechanism by which elevated FFA levels may impair insulin signaling is by its conversion to long-chain

fatty acyl-CoAs (LCFA-CoA) whose muscle levels have been shown to be negatively correlated with whole body insulin sensitivity⁴⁷. If not oxidized in the mitochondria, LCFA-CoAs are re-esterified via diacylglycerol (DAG) to form triglycerides and phospholipids. Increased intramyocellular triglyceride content is seen in obesity and type 2 diabetes⁴⁸ and is a strong predictor of insulin resistance in both animals and humans^{49,50}, probably as a marker of increased intracellular LCFA-CoAs and lipid intermediates. Mitochondrial function is also influenced by fat; the feeding of a high fat diet (50% fat) for 3 days to healthy volunteers resulted in the decreased expression of genes involved in oxidative phosphorylation in the mitochondria, as well as a decrease in the expression of PPAR γ coactivator1, consistent with mitochondrial dysfunction⁵¹ similarly, reduced mitochondrial oxidative phosphorylation activity could be demonstrated in the lean offspring of type 2 diabetic parents⁵². These findings suggest that the genes for mitochondrial biogenesis, while susceptible to environmental stimuli such as dietary intake of fat, may also be the genetic basis for the inheritance of diabetes.

While FFA promote a state of IR, studies have shown PUFA (particularly of n-3 origin) to exert protective action and hence, the dietary intake of an appropriate ratio of n-3 and n-6 fatty acids is important. Polyunsaturated n-3 fatty acids can act as agonists of peroxisome proliferation activation receptor (PPAR) γ ⁵³, similar to the therapeutic agents like the glitazones (which belong to the thiazolidinedione group). PPAR has several effects on gene expression⁵⁴, and can therefore, n-3 fatty acids can take part in several distinct but interacting pathways in muscle, and may increase GLUT 4 transcription⁵³. Intake of polyunsaturated fatty acids (PUFA) of marine origin, namely eicosapentaenoic acid (EPA; 20:5 n-3) and docosahexaenoic acid (DHA, 22:6 n-3), have been shown to be hypolipidemic, exert prophylactic effects on cardiovascular disease, protect against insulin resistance and obesity in rodents fed high-fat diets, and reduce insulin response to glucose in healthy humans⁵⁵⁻⁵⁷. The effects of n-3 and n-6 ratios can also be seen through studies of whole body insulin sensitivity which correlated with the amount of long chain, unsaturated fatty acids (C20–C22) in membrane

phospholipids of human thigh muscle⁵⁸ and it is reasonable to think that the fatty acid composition of muscle cell membranes reflects dietary fat intake, as demonstrated in animal studies⁵⁹. The importance of these observations cannot be overstated. Studies of populations specifically prone to obesity and type 2 diabetes (such as Pima American Indians) show that they have much lower levels of n-3 fatty acids in their muscle membrane lipids than Caucasian counterparts⁶⁰. More importantly, dietary interventions have been shown to increase n-3 fatty acids in skeletal muscle membrane phospholipids of obese subjects⁶¹, and reducing the n-6:n-3 ratio of dietary fat (to at least 10) prevented sucrose induced insulin resistance in rats⁶². In India, the intake of n-3 fatty acids is particularly low, and DHA intakes are practically non-existent⁶³. The consumption of vanaspati, a fat rich in trans-fatty acids is of concern, as this also leads to insulin resistance in rats⁶⁴.

Protein, Amino Acids and Muscle

The dietary intake of amino acids is also relevant, since essential amino acids produce a rapid increase in plasma amino acid concentrations and can stimulate skeletal muscle protein synthesis to a greater extent than nonessential amino acids⁶⁵. Indeed, amino acids given alone can never be utilized efficiently, and the combined effect of essential amino acids along with carbohydrate is optimal. In the elderly, studies have shown that the daily supplementation of hydroxymethylbutyrate, arginine and lysine for 12 wk positively alters measurements of functionality, strength, fat-free mass, and protein synthesis⁶⁶. The effect of lysine is of interest, particularly in view of the daily requirement of 30 mg/kg/day⁶⁷ or higher in chronic undernutrition⁶⁸, for this amino acid (which is more than double the requirement laid out in the 1985 WHO/FAO/UNU expert consultation⁶⁹) and its limiting nature in cereal based diets. Lysine intakes are marginal in low socio-economic group Indians^{67,68} and there are several lines of reasoning to suggest that lysine intake may influence muscle function and that protein accretion may be taking place in chronically undernourished individuals who are simply exposed to a more complete diet, without the benefit of additional exercise. First, well nourished Indians

showed a trend toward improved muscle function with higher lysine intakes⁷⁰, as have elderly women⁶⁶. Second, the lysine requirement of undernourished subjects placed on perfectly adequate diets for one week was 50% higher than that of well-nourished subjects⁶⁸. Third, in similar undernourished subjects, the leucine balance (an indicator of whole body protein balance) at different lysine intakes after they were successfully treated for intestinal parasites was positive at high lysine intakes⁷¹. None of these subjects increased their physical activity during these experiments. Fourth, a recent study using compilations of data on weight, height, dietary energy and protein from different regions of China showed that lower protein intake was related to growth failure while energy was not⁷². While this study did not distinguish between animal and plant protein, nor was it able to assign a value to the effect of micronutrients that accompany protein, it broadly reached similar conclusions as other studies which assessed the effect of animal protein on growth⁷³. If protein quality is assessed through the use of the protein digestibility adjusted amino acid score (PDCAAS), and assuming lysine to be the first limiting amino acid in cereal protein, then it becomes evident, with the new higher daily lysine requirement, that the protein quality of Indian diets is particularly poor with PDCAAS adjusted Protein:Energy (PE) ratios of about 0.08 for average Indian diets⁷⁴. This would mean that between 15 to 45% (depending on age and gender) of the sedentary Indian population would be at risk for protein deficiency, since a sedentary activity pattern would automatically raise the required PE ratio of the dietary requirement. This in itself is good cause to recommend increased physical activity in Indians, as it would then drop the PE ratio of their required dietary intake. Leucine is also an amino acid of interest as it can act as a signal to increase muscle protein synthesis in addition to its role as a substrate. However, this increased protein synthesis (in infusion protocols) shows tachyphylaxis in 2 hours explaining why chronically elevated leucine delivery does not elevate muscle protein synthesis in clinical situations⁷⁵. The stimulation of muscle protein synthesis by leucine is associated with an elevation of signaling activity in the mammalian target of rapamycin (mTOR)/p70 ribosomal subunit pathway⁷⁶. It is however, im-

portant to separate out the effect of amino acid supplementation from other factors, like for example, physical activity, since the latter can completely obscure the amino acid effects.

Exercise and Muscle

Finally, the effect of exercise dominates the environmental effects on muscle, and there are several epidemiological studies that have shown a beneficial effect of exercise on diabetes⁷⁷⁻⁸². Exercise and muscle contraction acutely activates glucose transport independent of insulin, and is associated with later effects on insulin sensitivity⁸³. Aerobic exercise can improve insulin sensitivity⁸⁴ through defined biochemical mechanisms in muscle^{85,86}. Resistance exercise has also been shown to improve whole-body insulin sensitivity in healthy males, through the acute reduction of intramyocellular lipids, although the lipid content of muscle is regained during recovery from exercise⁸⁷. It is possible that a lowered muscle mass may have a role in increasing fat accumulation as muscle fat oxidation, linked to daily physical activity, is an important determinant of whole body fat oxidation⁸⁸. A study on the effects of endurance and resistance training in non obese young women showed that insulin sensitivity improved through intrinsic (with endurance training) and muscle mass based (with resistance training) mechanisms, even though there were no changes in total body, subcutaneous or visceral fat after 6 months of training¹¹. Further evidence for the importance of physical activity and muscle mass comes from intervention studies, where a high physical activity has been shown to increase fat free mass (probably muscle) and reduce fat mass by small but significant amounts⁸⁹. Lifestyle interventions including diet and physical activity have also been shown reduce the incidence of diabetes, even when weight did not change⁹⁰. A similar decline in the age adjusted risk for CHD, hypertension and diabetes with increasing leisure time physical activity has also been demonstrated for middle aged men and women⁹¹.

Conclusion

In India, it would seem that an unfortunate combination of all the foregoing factors that promote IR are in place with nutritional and lifestyle

transitions resulting in high fat but low n-3 fatty acid intakes^{63,92}, coupled with a low physical activity^{93,94}, and a low quality protein intake^{71,75}. Indeed, the low physical activity puts an added requirement on the diet of a higher PE ratio, which is difficult to meet with diets that already have low protein quality. It would seem important therefore, to focus efforts on improving muscle physiology and function in addition to reducing fat, in the prevention of NCD. These intertwining environmental effects would not only increase total body fat mass, but might also result in a relatively lower body muscle content with less functionality, ultimately resulting in insulin resistance and non-communicable chronic disease.

The authors are from St John's Research Institute, St John's National Academy of Health Sciences, Bangalore 560034

References

1. Gopalan C. Rising incidents of obesity and diabetes. NFI Bulletin 2006
2. Misra A, Vikram N. Insulin resistance syndrome (metabolic syndrome) and Asian Indians. Curr Sci 2002, 83:1483-1496.
3. Banerji MA, Faridi N, Atluri R, Chaiken RL, Lebovitz HE. Body composition, visceral fat, leptin, and insulin resistance in Asian Indian men. J Clin Endocrinol Metab 1999, 84:137-144.
4. Deurenberg-Yap M, Chew SK, Deurenberg P. Elevated body fat percentage and cardiovascular risks at low body mass index levels among Singaporean Chinese, Malays and Indians. Obes Rev 2002, 3:209-215.
5. Dudeja V, Misra A, Pandey RM, Devina G, Kumar G, Vikram NK. BMI does not accurately predict overweight in Asian Indians in northern India. Br J Nutr 2001, 86:105-112.
6. Chowdhury B, Lantz H, Sjoström L. Computed tomography-determined body composition in relation to cardiovascular risk factors in Indian and matched Swedish males. Metabolism 1996, 45:634-644.
7. Misra A, Sharma R, Pandey RM, Khanna N. Adverse profile of dietary nutrients, anthropometry and lipids in urban slum dwellers of northern India. Eur J Clin Nutr 2001, 55:727-734.
8. Dela F, Larsen JJ, Mikines KJ, Ploug T, Petersen LN, Galbo H. Insulin stimulated muscle glucose clearance in patients with NIDDM. Effects of one legged physical training. Diabetes 1995, 44:1010-1020.
9. Ebeling P, Bourey R, Koranyi L, Tuominen JA, Groop LC, Henriksson J, Mueckler M, Sovijarvi A, Koivisto VA. Mechanism of enhanced insulin sensitivity in athletes. Increased blood flow, muscle glucose transport protein (glut-4) concentrations and glycogen synthase activity. J Clin Invest 1993, 92:1623-1631.
10. Houmard JA, Egan PC, Neuffer PD, Friedman JE, Wheeler WS, Israel RG, Dohm GL. Elevated skeletal muscle glucose transporter levels in exer-

cise-trained middle aged men. Am J Physiol 1991, 261:E437-E443.

11. Poehlman ET, Dvorak RV, DeNino WF, Brochu M, Ades PA. Effects of resistance training and endurance training on insulin sensitivity in nonobese, young women: a controlled randomized trial. J Clin Endocrinol Metab 2000, 85:2463-2468.

12. Naidu AN, Rao NP. Body mass index: a measure of the nutritional status in Indian populations. Eur J Clin Nutr 1994, 48(Suppl 3):S131-140.

13. Lee RC, Wang Z, Heo M, Ross R, Janssen I, Heymsfield SB. Total body skeletal muscle mass: development and cross validation of anthropometric prediction models. Am J Clin Nutr 2000, 72:796-803.

14. Song M-Y, Kim J, Horlick M, Wang J, Pierson Jr RN, Moonseong H, Gallagher D. Prepubertal Asians have less limb skeletal muscle. J Appl Physiol 2002, 92: 2285-2291.

15. Kurpad AV, Regan MM, Raj T Vasudevan J, Kuriyan R, Gnanou J, Young VR. Lysine requirement of chronically undernourished adult Indian subjects, measured by the 24h indicator amino acid oxidation and balance technique. Am J Clin Nutr 2003, 77: 101-108.

16. Chandalia M, Abate N, Garg A, Stray-Gundersen J, Grundy SM. Relationship between generalized and upper body obesity to insulin resistance in Asian Indian men. J Clin Endocrinol Metab 1999, 84:2329-2335.

17. Raji A, Seely EW, Arky RA, Simonson DC. Body fat distribution and insulin resistance in healthy Asian Indians and Caucasians. J Clin Endocrinol Metab 20001, 86:5366-5371.

18. Olsen DB, Sacchetti M, Dela F, Ploug T, Saltin B. Glucose clearance is higher in arm than leg muscle in type 2 diabetes. J Physiol 2005, 565.2: 555-562.

19. Albu JB, Kovera AJ, Allen L, Wainwright M, Berk E, Raja-Khan N, Janumala I, Burkey B, Heshka S, Gallagher D. Independent association of insulin resistance with larger amounts of intermuscular adipose tissue and a greater acute insulin response to glucose in African American than in white non-diabetic women. Am J Clin Nutr 2005, 82:1210-7.

20. Boesch, C, Slotboom H, Hoppeler H, Kreis R. In vivo determination of intra-myocellular lipids in human muscle by means of localized 1H-MR-spectroscopy. Magn Reson Med 1997, 37:484-493.

21. Gallagher D, Kuznia P, Heshka P, Albu J, Heymsfield SB, Goodpaster B, Visser M, Harris TB. Adipose tissue in muscle: a novel depot similar in size to visceral adipose tissue. Am J Clin Nutr 2005;81:903-10

22. Vaz M, Hunsberger S, Diffey B. Prediction equations for handgrip strength in healthy Indian male and female subjects encompassing a wide age range. Annals Hum Biol 2002; 29: 131-141.

23. Rantanen T, Harris T, Leveille SG, Visser M, Foley D, Masaki K, Guralnik JM. Muscle strength and body mass index as predictors of mortality in initially healthy men. J Gerontol 2000, 55A:M168-M173.

24. Vaz M, Thangam S, Prabhu A, Shetty PS. Maximal voluntary contraction as a functional indicator of adult chronic undernutrition. Br J Nutr 1996, 76, 9-15.

25. Oberbach A, Bossenz Y, Lehmann S, Niebauer J, Adams V, Paschke R, Schon MR, Bluher M, Punkt K.

- Altered fiber distribution and fiber-specific glycolytic and oxidative enzyme activity in skeletal muscle of patients with type 2 diabetes. *Diabetes Care* 2006, 29:895–900.
26. Henriksson J. Muscle fuel selection: effect of exercise and training. *Proc Nutr Soc* 1995, 54:125-38.
27. Mansell PI, Macdonald IA. The effect of starvation on insulin-induced glucose disposal and thermogenesis in humans. *Metabolism* 1990, 39:502-510.
28. Fery F, Plat L, Balasse EO. Effect of fasting on the intracellular metabolic partition of intravenously infused glucose in humans. *Am J Physiol* 1999, 277: E815-E823.
29. Tsintzas K, Jewell K, Kamran M, Laithwaite D, Boonsong T, Littlewood J, Macdonald I, Bennett A. Differential regulation of metabolic genes in skeletal muscle during starvation and refeeding in humans. *J. Physiol.* DOI: 10.1113/jphysiol. 2006.109892.
30. Park J, Berggren JR, Hulver MW, Houmard JA, Hoffman EP. GPR14, GPD1, and GDF8 as potential network collaborators in weight loss induced improvements in insulin action in human skeletal muscle *Physiol Genomics* DOI:10.1152/physiolgenomics.00045.2006
31. Gonzalez-Cadavid NF, and Bhasin S. Role of myostatin in metabolism. *Curr Opin Clin Nutr Metab Care* 2004, 7:451-457.
32. Kelley D, Mandarino L. Fuel selection in human skeletal muscle in insulin resistance: a reexamination. *Diabetes* 2000, 49:677-683.
33. Elks ML. Fat oxidation and diabetes of obesity: the Randle hypothesis revisited. *Medical Hypotheses* 1990, 33:257-260.
34. Kelley DE. Skeletal muscle fat oxidation: timing and flexibility are everything. *J Clin Invest* 2005, 115:1699-1702.
35. Ukropcova B, McNeil M, Sereda O, de Jonge L, Xie H, Bray GA, Smith SR. Dynamic changes in fat oxidation in human primary myocytes mirror metabolic characteristics of the donor. *J Clin Invest* 2005, 115:1934-1941.
36. Kelley DE, Goodpaster B, Wing RR, Simoneau JA. Skeletal muscle fatty acid metabolism in association with insulin resistance, obesity, and weight loss. *Am J Physiol* 1999, 277:E1130-E1141.
37. Zurlo F, Lillioja S, Esposito-Del Puente A, Nyomba BL, Raz I, Saad MF, Swinburn BA, Knowler WC, Bogardus C, Ravussin E. Low ratio of fat to carbohydrate oxidation as predictor of weight gain: A study of 24-h RQ. *Am J Physiol* 1990, 259:E650-E6570.
38. Chang L and Karin M. Mammalian MAP kinase signalling cascades. *Nature* 2001, 410: 37-40.
39. Forouhi NG, Jenkinson G, Thomas EL, Mullick S, Mierisova S, Bhonsle U, McKeigue PM, Bell JD. Relation of triglyceride stores in skeletal muscle cells to central obesity and insulin sensitivity in European and South Asian men. *Diabetologia* 1999, 42:932-935.
40. Roden M, Price BT, Perseghin G, Petersen KF, Rothman DL, Cline GW, Shulman GI. Mechanism of free fatty acid-induced insulin resistance in humans. *J Clin Invest* 1996, 97:2859–2865.
41. Storz P, Dox H, Wernig A, Pfizenmaier K Muller G. Cross-talk mechanisms in the development of insulin resistance of skeletal muscle cells. Palmitate rather than tumour necrosis factor inhibits insulin-dependent protein kinase B (PKB)/Akt stimulation and glucose uptake. *Eur J Biochem* 1999, 266: 17-25.
42. Griffin ME, Marcucci MJ, Cline GW. Free fatty acid-induced insulin resistance is associated with activation of protein kinase C theta and alterations in the insulin signaling cascade. *Diabetes* 1999, 48:1270-1274.
43. Paz K, Hemi R, LeRoith D. A molecular basis for insulin resistance. Elevated serine/threonine phosphorylation of IRS-1 and IRS-2 inhibits their binding to the juxtamembrane region of the insulin receptor and impairs their ability to undergo insulin-induced tyrosine phosphorylation. *J Biol Chem* 1997, 272:29911-29918.
44. Hirosumi J, Tuncman G, Chang L. A central role for JNK in obesity and insulin resistance. *Nature* 2002, 420: 333-336.
45. Uysal KT, Wiesbrock SM, Marino MW. Protection from obesity induced insulin resistance in mice lacking TNF. *Nature* 1997, 389:610-614.
46. Aguirre V, Uchida T, Yenush L. The c-Jun NH2-terminal kinase promotes insulin resistance during association with insulin receptor substrate-1 and phosphorylation of Ser307. *J Biol Chem* 2000, 275: 9047-9054.
47. Ellis BA, Poynten A, Lowy AJ. Long-chain acyl-CoA esters as indicators of lipid metabolism and insulin sensitivity in rat and human muscle. *Am J Physiol* 2000, 279: E554-560.
48. Anderwald C, Bernroider E, Krssak M. Effect of insulin treatment in type 2 diabetic patients on intracellular lipid content in liver and skeletal muscle. *Diabetes* 2002, 51: 3025-3032.
49. Kraegen EW, Clark PW, Jenkins AB. Development of muscle insulin resistance after liver insulin resistance in high-fat-fed rats. *Diabetes* 1991, 40:1397-1403.
50. Pan DA, Lillioja S, Kriketos AD, Milner MR, Baur LA, Bogardus C, Jenkins AB, Storlien LH. Skeletal muscle triglyceride levels are inversely related to insulin action. *Diabetes* 1997, 46:983-988.
51. Sparks LM, Xie H, Koza RA, Mynatt R, Hulver MW, Bray GA, Smith SR. A high-fat diet coordinately downregulates genes required for mitochondrial oxidative phosphorylation in skeletal muscle. *Diabetes* 2005, 54:1926–1933.
52. Shulman GI. Unraveling the cellular mechanism of insulin resistance in humans: new insights from magnetic resonance spectroscopy. *Physiology* 2004, 19:183-190.
53. Klier SA, Sundseth SS, Jones SA, Brown PJ, Wisely GB, Koble CS, Devchand P, Wahli W, Willson TM, Lenhard JM, Lehmann JM. Fatty acids and eicosanoids regulate gene expression through direct interactions with peroxisome proliferator-activated receptors alpha and gamma. *Proc Natl Acad Sci USA*, 1997; 94:4318–4323.
54. Deckelbaum RJ, Worgall TS, Seo T. n-3 Fatty acids and gene expression. *Am J Clin Nutr* 2006, 83:S1520-1525S.
55. Ruzickova J, Rossmeisl M, Prazak T, Flachs P, Sponarova J, Veck M, Trzicka E, Bryhn M, Kopecky J. Omega-3 PUFA of marine origin limit diet-induced obesity in mice by reducing cellularity of adipose tissue. *Lipids* 2004, 39:1177–1185.
56. Ruxton CH, Reed SC, Simpson MJ, Millington KJ. The health benefits of omega-3 polyunsaturated fatty acids: a review of the evidence. *J Hum Nutr Diet* 2004, 17:449–459.
57. Delarue J, LeFoll C, Corporeau C, Lucas D. N-3 long chain polyunsaturated fatty acids: a nutritional tool to prevent insulin resistance associated to type 2 diabetes and obesity? *Reprod Nutr Dev* 2004, 44:289–299
58. Borkman M, Storlien LH, Pan DA, Jenkins AB, Chisholm DJ, Campbell LV. The relationship between insulin sensitivity and the fatty acid composition of phospholipids of skeletal muscle. *N Eng J Med* 1993; 328: 328–344.
59. Storlien LH, Jenkins AB, Chisholm DJ, Pascoe WS, Khouri S, Kraegen EW. Influence of dietary fat composition on development of insulin resistance in rats: relationship to muscle triglyceride and omega-3 fatty acids in muscle phospholipids. *Diabetes* 1991 40:280-289
60. Pan DA, Lillioja S, Milner MR, Kriketos AD, Baur LA, Bogardus C, Storlien LH. Skeletal muscle membrane lipid composition is related to adiposity and insulin action. *J Clin Invest* 1995; 96:2802–2808
61. Haugaard SB, Madsbad S, Hoy CE, Vaag A. Dietary intervention increases n-3 long-chain polyunsaturated fatty acids in skeletal muscle membrane phospholipids of obese subjects. Implications for insulin sensitivity. *Clin Endocrinol (Oxf)* 2006, 64:169-178.
62. Ghafoorunissa, Ibrahim A, Rajkumar L, Acharya V. Dietary (n-3) long chain polyunsaturated fatty acids prevent sucrose-induced insulin resistance in rats. *J Nutr.* 2005,135:2634-8
63. Dwarkanath P, Muthayya S, Mhaskar A, Thomas A, Vaz M, Kurpad A. Essential fatty acid consumption in pregnant South Indian women: Relation with birth outcome. *Proceedings of the International Society for Study of Fats and Lipids* 2006, page 185 (A).
64. Ibrahim A, Natrajan S, Ghafoorunissa R. Dietary trans-fatty acids alter adipocyte plasma membrane fatty acid composition and insulin sensitivity in rats. *Metabolism* 2005, 54: 240-246.
65. Tipton KD, Gurki BE, Matin S, Wolfe RR. Nonessential amino acids are not necessary to stimulate net muscle protein synthesis in healthy volunteers. *J Nutr Biochem* 1999, 10: 89–95.
66. Flakoll P, Sharp R, Baier S, Levenhagen D, Carr C, Nissen S. Effect of beta-hydroxy-beta-methylbutyrate, arginine, and lysine supplementation on strength, functionality, body composition, and protein metabolism in elderly women. *Nutrition* 2004, 20:445-451.
67. Kurpad AV, Raj T, El-Khoury AE, Beaumier L, Kuriyan R, Srivatsa A, Borgonha S, Selvaraj A, Regan M and Young VR. Lysine requirements of healthy adult Indian subjects, measured by an indicator amino acid balance technique. *Am J Clin Nutr* 2000, 73:900-907.
68. Kurpad AV, Regan MM, Raj T Vasudevan J, Kuriyan R, Gnanou J, Young VR. Lysine requirement of chronically undernourished adult Indian subjects, measured by the 24h indicator amino acid oxidation and balance technique. *Am J Clin Nutr* 2003, 77: 101-108.
69. FAO/WHO/UNU. Energy and protein requirements: Report of a joint FAO/WHO/UNU Expert consultation. WHO Technical Report No 724 Geneva: WHO, 1985.
70. Kurpad AV, Regan MM, Raj T, El-Khoury AE, Kuriyan R, Vaz M, Chandakudlu D, Veena GV, Borgonha S, Young VR. The lysine requirement of healthy adult Indian subjects on longer term feed-

ing and measured by the 24h indicator amino acid oxidation and balance technique. *Am J Clin Nutr* 2002, 76:400-412.

71. Kurpad AV, Regan MM, Nazareth D, Nagaraj S, Gnanou J, Young VR. Intestinal parasites increase the dietary lysine requirement in chronically undernourished adult Indian subjects. *Am J Clin Nutr* 2003, 78:1145-1151.

72. Jamison DT, Leslie J, Musgrove P. Malnutrition and dietary protein: evidence from China and international comparisons. *Food Nutr Bull* 2003, 24:145-154.

73. Graham GG, Creed FM, Maclean WC Jr, Kallman CH, Rabold J, Mellitis ED. Determinants of growth among poor children: nutrient intake-achieved growth relationships. *Am J Clin Nutr* 1981, 34:539-554.

74. Millward DJ, Jackson AA. Protein/energy ratios of current diets in developed and developing countries compared with a safe protein/energy ratio: implications for recommended protein and amino acid intakes. *Public Health Nutrition* 2003, 7:387-405.

75. Rennie MJ, Bohe J, Smith K, Wackerhage H, Greenhaff P. Branched-chain amino acids as fuels and anabolic signals in human muscle. *J Nutr*. 2006, 136:264S-268S.

76. Christopher J. Lynch. Role of leucine in the regulation of mTOR by amino acids: revelations from structure-activity studies. *J Nutr*. 2001, 131:861S-865S.

77. Hu FB, Sigal RJ, Rich-Edwards JW, Colditz GA, Solomon CG, Willett WC, Speizer FE, Manson JE. Walking compared with vigorous physical activity and risk of type 2 diabetes in women: a prospective study. *JAMA* 1999, 282: 1433-1439.

78. Weinstein AR, Sesso HD, Lee IM, Cook NR, Manson JE, Buring JE, Gaziano JM. Relationship of physical activity vs body mass index with type 2 diabetes in women. *JAMA* 2004, 292:1188-1194.

79. Folsom AR, Kushi LH, Hong CP. Physical activity and incident diabetes mellitus in postmenopausal women. *Am J Public Health* 2000, 90:134-138.

80. Lynch J, Helmrich SP, Lakka TA, Kaplan GA, Cohen RD, Salonen R, Salonen JT. Moderately intense physical activities and high levels of cardiorespiratory fitness reduce the risk of non-insulin-dependent diabetes mellitus in middle-aged men. *Arch Intern Med* 1996, 156:1307-1314.

81. Hu FB, Leitzmann MF, Stampfer MJ, Colditz GA, Willett WC, Rimm EB. Physical activity and television watching in relation to risk for type 2 diabetes mellitus in men. *Arch Intern Med* 2001, 161:1542-1548.

82. Helmrich SP, Ragland DR, Leung RW, and Paffenbarger RS Jr. Physical activity and reduced occurrence of non-insulin-dependent diabetes mellitus. *N Engl J Med* 1991, 325:147-152.

83. Holloszy JO. Exercise-induced increase in muscle insulin sensitivity. *J Appl Physiol* 2005, 99: 338-343.

84. Henriksson J. Influence of exercise on insulin sensitivity. *J Cardiovasc Risk* 1995, 2:303-309.

85. Henriksen EJ. Effects of acute exercise and exercise training on insulin resistance. *J Appl Physiol* 2002, 93:788-796.

86. Perseghin G, Price TB, Petersen KF, Roden M, Cline GW, Gerow K, Rothman DL, Shulman GI. Increased glucose transport-phosphorylation and muscle glycogen synthesis after exercise training

in insulin-resistant subjects. *N Engl J Med* 1996, 335:1357-1362.

87. Koopman R, Manders RJ, Jonkers RA, Hul GB, Kuipers H, van Loon LJ. Intramyocellular lipid and glycogen content are reduced following resistance exercise in untrained healthy males. *Eur J Appl Physiol*. 2006, 96:525-534.

88. Morio B, Hocquette J-F, Montaurier C, Boirie Y, Bouteloup-Demange C, McCormack C, Fellmann N, Beaufriere B, Ritz P. Muscle fatty acid oxidative capacity is a determinant of whole body fat oxidation in elderly people. *Am J Physiol* 2001, 280: E143-E149.

89. Wilmore JH, Despres J-P, Stanforth PR, Mandel S, Rice T, Gagnon J, Leon AS, Rao DC, Skinner JS, Bouchard C. Alterations in body weight and composition consequent to 20 wk of endurance training: the HERITAGE family study. *Am J Clin Nutr* 1999, 70:346-352.

90. Tuomilehto J, Lindstrom J, Eriksson JG, Valle TT, Hamalainen H, Ilanne-Parikka P, Keinanen-Kiukkaanniemi S, Laakso M, Louheranta A, Rastas M, Salminen V, Uusitupa M; Finnish Diabetes Prevention Study Group. Prevention of type 2 diabetes mellitus by changes in lifestyle among subjects with impaired glucose tolerance. *N Engl J Med* 2001, 344:1343-1350.

91. Haapanen N, Miilunpalo S, Vuori I, Oja P, Pasanen M. Association of leisure time physical activity with the risk of coronary heart disease, hypertension and diabetes in middle aged men and women. *Int J Epidemiol* 1997, 26:739-747.

92. Ghafoorunissa. Requirements of dietary fats to meet nutritional needs and prevent the risk of atherosclerosis. *In J Med Res* 1998, 108:191-202.

93. Borgonha S, Shetty PS, Kurpad AV. Total energy expenditure and physical activity in chronically undernourished Indian males measured by the doubly labeled water method. *Ind J Med Res* 2000, 111: 24-32.

94. Vaz M, Yusuf S, Bharath AV, Kurpad AV, Swaminathan S. The nutrition transition in India. *South Afr J Clin Nutr* 2005, 18:198-201.

FOUNDATION NEWS

● **The Annual Day of the Foundation** was celebrated on November 29, 2006.

C Ramachandran Memorial Lecture

Dr D Banerji, Professor Emeritus, Department of Social Medicine and Community Health, JNU delivered the C Ramachandran Memorial Lecture on "Primary Health Care in India: How to achieve what Bhore Committee envisaged?"

Symposium on "Primary Health Care – New Initiatives"

NFI in collaboration with Govern-

ment of India and WHO organised a two-day symposium on "Primary Health Care – New Initiatives" at India International Centre, on November 30th and December 1st, 2006. The Symposium was inaugurated by Mr Cecilio Adorna (Country Representative, UNICEF).

● **National Academy of Medical Sciences (NAMS)**

In recognition of his outstanding contribution to Medical Sciences and Biochemical Research, Dr C Gopalan – President NFI, has been conferred Life Time Achievement Award of NAMS for the year 2006.

Dr Prema Ramachandran, Director NFI has been elected as the President of NAMS. She will hold the office for three years.

NUTRITION NEWS

● **Nutrition Society of India**

The XXXVIII Annual Conference of Nutrition Society of India was held in November 2006 at AIH&PH, Kolkata. The theme of the meeting was "Nutrition in Different Stages". Dr. M.K. Bhan, delivered the thirtieth Gopalan Oration on "Preparing to face the Challenge". Eighteenth Srikantia Memorial Lecture was delivered by Dr. K. Vijayaraghavan, on "Community Nutrition Research in India – Contributions, Constraints and Controversies". During the Conference two symposia were also organised: "Infant Feeding and Growth" and "Nutrition and Development"

Mrs Anshu Sharma from NFI presented a paper on "Detection and Management of Anaemia in Pregnancy in an Urban Primary Health Care Institution" in this meeting and was awarded Young Scientists' Senior Award in Community Nutrition.

● **Kamla Puri Sabharwal Memorial Lecture**

Dr Sushma Palmer delivered the XXXIII Kamla Puri Sabharwal Memorial Lecture on November 16, 2006. The title of the lecture was "Global Health in Transition: The case for eating Green and Promoting Health".